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Abstract: Sunlamp irradiation of 1-substituted 5-iodo-1-pentynes, 5 equiv of phenyl isocyanide, and 1.5 equiv of hexamethylditin in *tert*-butylbenzene (0.01–0.025 M) at 150 °C produces 9-substituted 2,3-dihydro-1*H*-cyclopenta[b]quinolines in 36–70% yields. A mechanistic proposal for this first example of a 4 + 1 radical annulation includes the following: (1) radical addition to an isonitrile, (2) cyclization of the resulting imidoyl radical to the alkyne, (3) addition of the so-formed vinyl radical to the aromatic ring, and (4) rearomatization. When substituted isonitriles (*p*-F, *p*-OMe, *m*-F) are employed, the major unrearranged products are accompanied by 7–30% of rearranged products. Independent generation of a proposed intermediate suggests that the rearranged products arise by initial closure of the vinyl radical to form a five-membered ring, followed by C–N bond cleavage and recyclization.

Introduction

The sequencing of radical reactions is emerging as a powerful strategy for the rapid construction of diverse classes of molecules.² Radical annulations, which sequence an addition reaction before a cyclization, provide direct entries to functionalized five-membered rings from acyclic precursors.³ As Figure 1 illustrates, all existing radical annulations are of the 3 + 2 (or, more recently, $4 + 2^{3i,k}$) class.⁴ While a number of reagents have been used as equivalents of the synthon 1, an alkene 3 is invariably used as the two-atom component because it is naturally suited as a vicinal radical acceptor/radical donor (see synthon 2). The lower half of Figure 1 introduces a strategy for conducting 4 + 1 annulations that combines synthon 4 with a geminal radical acceptor/radical donor 5. Literature precedent⁵⁻⁷ indicates that either an isonitrile 6 or carbon monoxide^{8,9} (7) should serve as the actual reagent

(4) In this new retrosynthetic notation, closed dots (•) represent radical doner sites and open dots (•) represent radical acceptor sites. See ref 2b and Curran, D. P. SynLett 1991, 63.

(5) Examples of radical additions to isonitriles: (a) Saegusa, T.; Kobayashi, S.; Ito, Y.; Yasuda, N. J. Am. Chem. Soc. **1968**, 90, 4182. (b) Meier, M.; Rüchardt, C. Tetrahedron Lett. **1983**, 24, 4671. (c) Stork, G.; Sher, M. M. J. Am. Chem. Soc. **1983**, 105, 6765. (d) Barton, D. H. R.; Ozbalik, N.; Vacher, B. Tetrahedron **1988**, 44, 3501.

(6) Structure of imidoyl radicals: (a) Blum, P. M.; Roberts, B. P. J. Chem. Soc., Perkin Trans. 2 1978, 1313. (b) Davies, A. G.; Nedelec, J.-Y.; Sutcliffe, R. J. Chem. Soc., Perkin Trans. 2 1983, 209.

(7) Additions and cyclizations of imidoyl radicals: (a) Leardini, R.; Pedulli, G. F.; Tundo, A.; Zanardi, G. J. Chem. Soc., Chem. Commun. 1984, 1320. (b) Leardini, R.; Nanni, D.; Pedulli, G. F.; Tundo, A.; Zanardi, G. J. Chem. Soc., Perkin Trans. 1 1986, 1591. (c) Leardini, R.; Nanni, D.; Tundo, A.; Zanardi, G. J. Chem. Soc., Chem. Commun. 1989, 757. (d) Leardini, R.; Nanni, D.; Tundo, A.; Zanardi, G. Gazz. Chem. Ital. 1989, 119, 637. (e) Bachi, M.; Denenmark, D. J. Am. Chem. Soc. 1989, 111, 1886. (f) Bachi, M.; Denenmark, D. J. Org. Chem. 1990, 55, 3442.

(8) Leading reference for additions to CO: Ryu, 1; Kusano, K.; Ogawa, A.; Kambe, N.; Sonoda, N. J. Am. Chem. Soc. 1990, 112, 1295.

equivalent of synthon 5. We now describe a one-step synthesis of cyclopentaquinolines from isonitriles. This is the first example of a 4 + 1 radical annulation, and it illustrates that it should be possible to develop a whole class of n + 1 radical annulations.

Results and Discussion

Preliminary experiments conducted by heating iodopentyne (8a), phenyl isocyanide (9), and tributyltin hydride in benzene at 80 °C were not encouraging (Figure 2). However, we were able to isolate small amounts of a highly UV-active product that we quickly identified as the known quinoline 10a.¹⁰ Despite the presence of tin hydride, 10a is not a reduction product, but instead it maintains the oxidation state of the starting materials. By individually changing the reaction variables (temperature, concentration, stoichiometry, added reagents), we identified conditions that produced useful amounts of 10a. From these experiments, we learned the following: (1) The concentration of the reaction is very important. Higher yields are obtained at lower concentrations. (2) The reaction is highly temperature sensitive. Temperatures well above 80 °C are required to produce significant vields of 10a. (3) Excess isonitrile is required, probably because it suffers thermal decomposition during the reaction.

When a mixture of 1 equiv of 8a (0.025 M), 5 equiv of phenyl isocyanide, 1.5 equiv of tributyltin hydride, and 0.25 equiv of azobisisobutyronitrile (AIBN) was heated at 150 °C in a sealed tube in benzene for 6 h, iodide 8a was consumed and we isolated 10a in 30% yield after flash chromatography. An analysis of a crude reaction in C_6D_6 indicated that a large amount (>30%) of 1-pentyne also formed. This volatile product results from reduction of the pentynyl radical by tin hydride. We isolated much better yields of 10a when we substituted hexamethylditin for tributyltin hydride, provided that we irradiated the heated reaction with a sunlamp. Under a standard set of conditions, sunlamp irradiation of a tert-butylbenzene solution (150 °C) of 8a (0.025 M), 9 (5 equiv), and hexamethylditin (1.5 equiv) for 38 h gave 10a in 63% isolated yield. Little or no 1-pentyne formed in this reaction, and no other low molecular weight products were evident by ¹H NMR analysis or by chromatography. However, there was a significant

(9) We have conducted a preliminary experiment that shows that annulations with CO should be possible.



(10) Borsche, W. Ann. 1910, 377, 70.

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Table I. Cyclopentaquinolines from 4 + 1 Annulations



3+2 Radical Annulations



4 + 1 Radical Annulations



Figure 1.



Figure 2.

amount of polar, high molecular weight material that probably resulted from thermal decomposition of the isonitrile.

We surveyed the generality of this procedure by reacting several iodopentynes with phenyl isocyanide (9), p-fluorophenyl isocyanide (11), p-methoxyphenyl isocyanide (14), and m-fluorophenyl isocyanide (17). Table I summarizes the results of this survey. All of the products in this table were isolated in pure form, and their structures were identified by standard spectroscopic techniques.¹¹

For identifying regioisomers (especially in entries 10 and 11), ¹⁹F couplings to ¹H and ¹³C were useful, as were nuclear Overhauser effect (NOE) experiments. The Experimental Section contains pertinent results from these experiments. When heated with phenyl isocyanide, terminally substituted iodopentynes 8b-d produced 9-substituted cyclopenta-fused quinolines 10b-d (entries 1-3). With p-fluorophenyl isocyanide (11), two separable regioisomers (12 and 13) formed in ratios that varied slightly as a function of the alkyne substituent (entries 4-6). The minor regioisomer 13 is a rearranged product-the orientation of the C-N bond and the fluorine has changed from para to meta. Qualitatively similar results were obtained with *p*-methoxyphenyl isocyanide (14); the major product 15 was accompanied by lesser amounts of the rearranged product 16 (entries 7-9). Finally, the use of m-fluorophenyl isocyanide (17) gave rise to a mixture of four products (entries 10 and 11): two unrearranged (18 and 13), and two rearranged (12 and 19). Again, the unrearranged products predominated. For each of the three isonitriles, the same trend was observed; the amount of rearranged product decreased in the series 8b (R = Me) > 8a (R = H) > 8d (R = Ph).

This new quinoline synthesis consists of a fascinating sequence of radical reactions that is dissected in eqs 1-13. The 4 + 1 radical annulation is an integral part of this sequence. We envision that radical **20** may be formed either by photolytic cleavage of the ditin, followed by iodine abstraction (eq 1) or by direct photolytic cleavage of the C-I bond (eq 2).¹² In the latter case, the ditin

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10d: Fehnel, E. A. J. Org. Chem. 1966, 31, 2899.
15a and 16a: Kazoko, O.; Hiroko, S.; Yujiro, N. Nippon Kagaku Kaishi 1989, 846 (Chem. Abstr. 1989, 111, 173960b).
12a: Yen, V. Q.; Buu-Hoi, N. P.; Xuong, N. D. J. Org. Chem.
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scavenges atomic (or molecular) iodine. Radical generation is followed by three successive, precedented carbon-carbon-forming reactions: addition of an alkyl radical to an isonitrile,⁵ cyclization of an imidoyl radical to an alkyne,⁷ and cyclization of a vinyl radical to a phenyl ring.¹³ Ultimately, an oxidation is required to form the final product.

Addition of 20 to the isonitrile gives radical 21 (eq 3). Like vinyl and acyl radicals, such imidoyl radicals are bent rather than linear⁶ and the structure of the product dictates that cyclization must occur predominantly in a 5-exo fashion to form 22Z (eq 4).



There are no confirmed examples of 5-exo ring openings of vinyl radicals, so we believe that this cyclization is irreversible. We do not know whether 21E is formed kinetically on addition and it cyclizes more rapidly than it interconverts with 21Z or whether 21E and 21Z are in rapid equilibrium and cyclization occurs through 21E (Curtin-Hammett kinetics).¹⁴ Related disubstituted vinyl radicals have very low inversion barriers (<5 kcal/mol),¹⁵ and cyclizations cannot usually compete with isomerizations. Compared to vinyl radicals, imidoyl radicals should have higher inversion barriers¹⁶ (the electronegative nitrogen pyramidalizes the radical). But it is by no means clear that these barriers are sufficiently high so that, if **21***E* were formed kinetically, it could not isomerize to 21Z. To further cloud the isomerization picture, we cannot rule out the possibility that 21Z does cyclize to some extent but that the product radical decomposes by nonproductive pathways because further cyclization to the aromatic ring is geometrically prohibited. To understand the details of this cy-

clization, we need a better understanding of the factors that control equilibria and rates of reaction of stereoisomeric sp²-hybridized radicals.

Cyclic vinyl radical 22 is certainly rapidly inverting,¹⁵ and the depicted isomer can cyclize to either of two positions on the aromatic ring (eqs 5 and 6). Closure to the ortho position forms six-membered-ring product 23, which is then ultimately converted to the unrearranged product 24 (eq 6). Rearranged product 26



must ultimately arise from cyclization of 22 to the ipso position to form five-membered-ring product 25 (eq 5). That such vinyl radicals can close to either the ipso or the ortho position is a rather general observation,^{7,17} and although ipso product 25 may lead to 24 as well as 26, there is some evidence that it does not.7b

We envision at least two ways (eqs 7 and 8) by which radical 25 can isomerize to 28, the probable precursor of rearranged product 26. The first possibility is that 25 may suffer ring opening.



Reverse cyclization to reform vinyl radical 22 is unlikely, but ring opening of 25 with C-N cleavage to give 27 is a reasonable possibility (eq 7). Closure of iminyl radical 27 at the ortho position gives the rearranged radical 28. Newcomb and co-workers have recently shown that aminyl radical cyclizations are surprisingly easily reversed,¹⁸ and this is the closest current precedent for the reverse cyclization of radical 25 to give 27. The second possibility is that radical 25 undergoes a (reversible?) 3-exo closure to give strained intermediate 29 (eq 8). 3-Exo opening of 29 by cleavage of the intraannular bond then gives 28. Such ring expansions are well precedented for simple β multiply bonded alkyl radicals,¹⁹ although we know of no examples where allyl or dienyl radicals participate.

A simple experiment proved that if 27 were formed, it would ultimately be converted to 28 under the reaction conditions (eq 9). Cyclization of bromide 30a under the standard conditions provided a 6/1 mixture of quinoline 10d and the reduced product. Quinoline 10d was isolated in 68% yield. It is highly probable that the vinyl radical derived from bromine abstraction of 30 cyclizes to form 27 (R = Ph, X = H). Thus, 27 is a viable, but not obligatory, intermediate in the conversion of 25 to 28 (eq 7).

Cyclization of the *p*-fluoro derivative 30b provided a regiochemical marker: 6-fluoro isomer 31 was the only product formed, and it was isolated in 76% yield. Thus, if radical 27 is formed in the radical annulation, it leads only to the rearranged product

⁽¹³⁾ For related cyclizations of alkyl and vinyl radicals to make quinoline derivatives, see ref 7

⁽¹⁴⁾ Seeman, J. 1. Chem. Rev. 1983, 83, 83.

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⁽¹⁶⁾ INDO calculations on •CH=NH predict that the E isomer is more stable than the Z isomer by ~ 2.5 kcal/mol and that the barrier to interconversion is ~ 10 kcal/mol. See ref 6a.

⁽¹⁷⁾ For related ipso closures of aryl radicals, see: (a) Snieckus, V.; Cuevas, J.-C.; Sloan, C. P.; Liu, H.; Curran, D. P. J. Am. Chem. Soc. 1990, 112, 896.
 (b) Grimshaw, J.; Haslett, R. J. J. Chem. Soc., Perkin Trans. 1
 1980, 657.
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30a Ar = C_6H_5 30b Ar = *p*-FC_6H_4



26 and not the unrearranged product 24.

Accepting that radicals 23 and 28 are the precursors of the unrearranged 24 and rearranged 26 products, then the last step must be an oxidation (eq 10). Even under reductive conditions,

$$(10)$$

23• + Me₃SnI $\xrightarrow{\text{electron}}$ 24 + Me₃Sn• + HI (11) transfer

HI + Me₃SnSnMe₃ \longrightarrow Me₃SnI + Me₃SnH (12) 23. + 23. \longrightarrow H_{1} H_{2} H_{2}

the formation of rearomatized products after radical additions to aromatic rings is $common;^{17,20}$ however, the oxidant is not immediately obvious. One possibility is that oxygen is present in trace amounts. Although we do not rigorously remove oxygen by freeze/thaw techniques, our reactions are degassed, and we do not believe that sufficient oxygen remains to conduct stoichiometric oxidations. The possibility that the starting iodide is an oxidant can be discarded because this would result in a chain,²¹ and stoichiometric quantities of tin reagents would not be required. A second, related possibility is that trimethyltin iodide is the oxidant (eq 11). This would also result in a chain, but the tin reagent would be consumed as a reagent because the HI formed on deprotonation of 30 would react with the hexamethylditin to form trimethyltin iodide and trimethyltin hydride (eq 12). If this chain is occurring, we do not believe that it is very efficient. The high temperature and long time requirements make efficient chains seem unlikely, and we have no evidence that trimethyltin hydride accumulates as a reaction product. A final possibility is that oxidation occurs by radical disproportionation (eq 13). Disproportionation of two molecules of 23 would give 24 and dihydroquinoline 33 or a double bond isomer. Since the yield of quinoline often exceeds 50%, we must then postulate that the dihydroquinolines are air oxidized to 24 during the reaction or workup. While this oxidation is very likely,²² there is no evidence for dihydroquinoline products when reactions conducted in C_6D_6 are subjected to NMR analysis prior to exposure to air.

In conclusion, the reactions of iodopentynes with aryl isonitriles provide a convenient, one-step synthesis of cyclopenta-fused isoquinolines. The success of these reactions raises a variety of interesting mechanistic and structural questions about the intermediate radicals. Of more immediate importance, it demonstrates that 4 + 1 radical annulations of isonitriles are feasible and variations on this general theme are almost certainly possible. For example, by an appropriate choice of substituents and reactions conditions, it should be possible to intercept the radical 22 directly after the 4 + 1 annulation (steps 3 and 4). This would provide a new route to imine derivative of cyclopentanones.

Experimental Section

General Procedure. To a flat flask (Pyrex, outside dimensions 36 × 160×8 mm, wall thickness 1 mm) under N₂ was added a 1-substituted-5-iodo-1-pentyne (0.25 mmol), an isonitrile (1.25 mmol), hexamethylditin (0.375 mmol), and tert-butylbenzene (10 mL). The mixture was irradiated with a 275-W sunlamp (GE RSK6) at 150 °C (silicone oil bath, about two-thirds of the liquid body higher than the oil level) for 12 to 60 h until the iodopentyne was consumed (followed by ¹H NMR). The reaction was then cooled to room temperture, diluted with ether (15 mL), shaken with 2 N HCl (10 mL), filtered through a sintered funnel, and separated. The organic layer was extracted with 2 N HCl (10 mL \times 4). The combined aqueous phase was neutralized with 6 N NaOH and was extracted with ether (30 mL \times 3). The combined ether extract was washed successively with aqueous NaHCO₃, H₂O, and brine (30 mL each) and dried over MgSO₄. After solvent removal, the residue was purified by chromatography (aluminum oxide 90, EM product) to provide the 9-substituted-2,3-dihydro-1H-cyclopenta[b]quinoline isomers. If needed, further separation was accomplished by MPLC or HPLC.

2,3-Dihydro-1*H***-cyclopenta**[*b*]**quino**line (10a): ¹H NMR (300 MHz, CDCl₃) δ 8.02 (1 H, d, J = 8.5 Hz), 7.90 (1 H, br s), 7.74 (1 H, dd, J = 8.1, 1.3 Hz), 7.62 (1 H, td, J = 7.7, 1.3 Hz), 7.45 (1 H, td, J = 7.5, 1.0 Hz), 3.17 (2 H, t, J = 7.7 Hz), 3.10 (2 H, td, J = 7.5, 1.3 Hz), 2.22 (2 H, qt, J = 7.6 Hz); ¹³C NMR (75 MHz, CDCl₃) δ 167.54, 147.19, 135.26, 129.98, 128.24, 128.04, 127.20, 127.11, 125.20, 34.32, 30.18, 23.29; IR (neat) 3062, 2963, 1502, 1408, 756 cm⁻¹; MS *m/e* 169 (M), 168 (M - 1, base peak); HRMS for C₁₂H₁₁N calcd 169.0891, found 169.0891.

9-Methyl-2,3-dihydro-1*H*-cyclopenta[*b*]quinoline (10b): ¹H NMR (300 MHz, CDCl₃) δ 8.01 (1 H, dd, *J* = 8.3, 1.0 Hz), 7.94 (1 H, dd, *J* = 8.3, 1.2 Hz), 7.61 (1 H, td, *J* = 7.6, 1.2 Hz), 7.48 (1 H, td, *J* = 7.7, 1.0 Hz), 3.17 (2 H, t, *J* = 7.7 Hz), 3.06 (2 H, t, *J* = 7.4 Hz), 2.59 (3 H, s), 2.20 (2 H, qt, *J* = 7.6 Hz); ¹³C NMR (75 MHz, CDCl₃) δ 167.02, 147.49, 138.07, 134.03, 129.21, 128.00, 127.07, 125.23, 123.37, 35.16, 29.66, 23.00, 14.91; IR (neat) 3063, 2955, 1613, 1570, 1507, 1389, 1340, 1215, 1024, 758 cm⁻¹; MS *m/e* 184 (M + 1), 183 (M, base peak), 182 (M - 1), 168; HRMS for C₁₃H₁₃N calcd 183.1048, found 183.1048.

9-Benzyl-2,3-dihydro-1*H*-cyclopenta[*b*]quinoline (10c): ¹H NMR (300 MHz, CDCl₃) δ 8.03 (1 H, dd, J = 7.7, 1.0 Hz), 7.92 (1 H, dd, J = 7.7, 1.2 Hz), 7.60 (1 H, td, J = 7.7, 1.2 Hz), 7.42 (1 H, td, J = 7.7, 1.0 Hz), 7.24–7.15 (3 H, m), 7.10 (2 H, d, J = 7.6 Hz), 4.41 (2 H, S), 3.21 (2 H, t, J = 7.7 Hz), 3.05 (2 H, t, J = 7.4 Hz), 2.21 (2 H, qt, J = 7.6 Hz); ¹³C NMR (75 MHz, CDCl₃) δ 167.44, 148.13, 139.94, 138.72, 135.00, 129.37, 128.66 (2 C), 128.24 (2 C), 128.12, 126.66, 126.36, 125.62, 123.95, 35.16, 34.87, 29.77, 23.07; IR (neat) 3072, 2957, 1611, 1577, 1498, 754, 705 cm⁻¹; MS *m/e* 260 (M + 1), 259 (M, base peak), 258 (M – 1), 181, 168, 91.77; HRMS for C₁₉H₁₇N calcd 259.1361, found 259.1361.

9-Phenyl-2,3-dihydro-1*H*-cyclopenta[*b*]quinoline (10d): ¹H NMR (300 MHz, CDCl₃) δ 8.07 (1 H, dd, *J* = 8.8, 0.8 Hz), 7.64–7.59 (2 H, m), 7.56–7.43 (3 H, m), 7.40–7.35 (3 H, m), 3.24 (2 H, t, *J* = 7.6 Hz), 2.91 (2 H, t, *J* = 7.4 Hz), 2.17 (2 H, qt, *J* = 7.5 Hz); ¹³C NMR (75 MHz, CDCl₂) δ 167.40, 147.94, 142.63, 136.72, 133.61, 129.26 (2 C), 128.80, 128.47 (2 C), 128.19, 127.95, 126.17, 125.62, 125.46, 35.20, 30.31, 23.50; IR (neat) 3064, 2973, 1577, 1487, 767, 759, 696 cm⁻¹; MS *m*/*e* 246 (M + 1), 245 (M, base peak), 244 (M – 1), 168; HRMS for C₁₈H₁₅N caled 245.1024, found 245.1024.

6-Fluoro-2,3-dihydro-1*H*-cyclopenta[*b*]quinoline (13a): ¹H NMR (300 MHz, CDCl₃) δ 7.88 (1 H, s), 7.71 (1 H, dd, *J* = 8.9, 6.1 Hz), 7.64 (1 H, dd, *J* = 10.4, 2.6 Hz), 7.25 (1 H, td, *J* = 8.6, 2.6 Hz), 3.15 (2 H, t, *J* = 7.7 Hz), 3.08 (2 H, t, *J* = 7.4 Hz), 2.22 (2 H, qt, *J* = 7.5 Hz); ¹³C NMR (75 MHz, CDCl₃) δ 169.22, 162.53 (*J*_{CF} = 248.9 Hz), 148.44 (*J*_{CF} = 8.1 Hz), 135.14, 130.32, 129.21 (*J*_{CF} = 9.7 Hz), 124.38, 115.75

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⁽²²⁾ Francis, R. F.; Crews, C. D.; Scott, B. S. J. Org. Chem. 1978, 43, 3227.

 $(J_{CF} = 24.4 \text{ Hz})$, 112.48 $(J_{CF} = 19.6 \text{ Hz})$, 34.72, 30.45, 23.68; IR (neat) 3038, 2963, 1632, 1505, 1414, 1223, 911 cm⁻¹; MS *m/e* 188 (M + 1), 187 (M), 186 (M - 1, base peak); HRMS for C₁₂H₁₀FN calcd 187.0797, found 187.0770.

8-Fluoro-2,3-dihydro-1*H*-cyclopenta[*b*]quinoline (18a): ¹H NMR (500 MHz, CDCl₃) δ 8.17 (1 H, s), 7.81 (1 H, d, *J* = 8.5 Hz), 7.53 (1 H, td, *J* = 8.1, 6.1 Hz), 7.13 (1 H, dd, *J* = 9.6, 8.1 Hz), 3.17 (2 H, t, *J* = 7.7 Hz), 3.12 (2 H, td, *J* = 7.5, 1.2 Hz), 2.23 (2 H, qt, *J* = 7.6 Hz); ¹³C NMR (125 MHz, CDCl₃) δ 169.06, 158.153 (*J*_{CF} = 253.0 Hz), 148.44, 136.10, 127.86 (*J*_{CF} = 9.0 Hz), 124.40, 123.50, 118.01 (*J*_{CF} = 16.2 Hz), 109.44 (*J*_{CF} = 19.7 Hz), 34.74, 30.73, 23.62; IR (neat) 3033, 2971, 1507, 1188, 810, 758 cm⁻¹; MS *m/e* 188 (M + 1), 187 (M), 186 (M - 1, base peak); HRMS for C₁₂ for C₁₂H₁₀FN calcd 187.0797, found 187.0797.

5-Fluoro-2,3-dihydro-1*H*-cyclopenta[*b*]quinoline (19a): ¹H NMR (500 MHz, CDCl₃) δ 7.91 (1 H, s), 7.52 (1 H, d, *J* = 8.1 Hz), 7.38 (1 H, td, *J* = 7.9, 5.0 Hz), 7.32 (1 H, ddd, *J* = 10.8, 7.7, 1.3 Hz), 3.22 (2 H, t, *J* = 7.7 Hz), 3.11 (2 H, td, *J* = 7.5, 1.2 Hz), 2.23 (2 H, qt, *J* = 7.6 Hz); ¹³C NMR (125 MHz, CDCl₃) δ 168.58, 157.92 (*J*_{CF} = 254.4 Hz), 137.54 (*J*_{CF} = 12.4 Hz), 137.04, 130.23, 129.26, 125.30 (*J*_{CF} = 7.8 Hz), 123.12 (*J*_{CF} = 4.8 Hz), 112.70 (*J*_{CF} = 19.4 Hz), 34.82, 30.63, 23.67; IR (neat) 3031, 2953, 1496, 1250, 1125, 1026, 893, 762 cm⁻¹; MS *m*/*e* 188 (M + 1), 187 (M, base peak), 186 (M - 1); HRMS for C₁₂H₁₀FN calcd 187.0797, found 187.0779.

7-Fluoro-9-methyl-2,3-dihydro-1*H*-cyclopenta[*b*]quinoline (12b): ¹H NMR (300 MHz, CDCl₃) δ 7.98 (1 H, dd, *J* = 9.1, 5.7 Hz), 7.53 (1 H, dd, *J* = 10.3, 2.7 Hz), 7.37 (1 H, td, *J* = 8.6, 2.7 Hz), 3.15 (2 H, t, *J* = 7.7 Hz), 3.06 (2 H, t, *J* = 7.4 Hz), 2.54 (3 H, s), 2.21 (2 H, qt, *J* = 7.6 Hz); ¹³C NMR (125 MHz, CDCl₃) δ 166.56, 160.10 (*J*_{CF} = 244.5 Hz), 144.55, 137.53 (*J*_{CF} = 4.9 Hz), 134.91, 131.37 (*J*_{CF} = 9.3 Hz), 127.93 (*J*_{CF} = 9.1 Hz), 117.74 (*J*_{CF} = 25.3 Hz), 107.23 (*J*_{CF} = 2.6 Hz); MS *m*_ℓ 202 (M + 1), 201 (M, base peak), 200 (M - 1), 186; HRMS for C₁₃H₁₂FN calcd 201.0954, found 201.0954.

6-Fluoro-9-methyl-2,3-dihydro-1*H*-cyclopenta[*b*]quinoline (13b): ¹H NMR (300 MHz, CDCl₃) δ 7.92 (1 H, dd, *J* = 9.2, 6.1 Hz), 7.63 (1 H, dd, *J* = 10.3, 2.6 Hz), 7.26 (1 H, td, *J* = 8.7, 2.6 Hz), 3.16 (2 H, t, *J* = 7.7 Hz), 3.05 (2 H, t, *J* = 7.5 Hz), 2.58 (3 H, s), 2.21 (2 H, qt, *J* = 7.6 Hz); ¹³C NMR (75 MHz, CDCl₃) δ 168.44, 162.36 (*J*_{CF} = 249.5 Hz), 148.57 (*J*_{CF} = 12.2 Hz), 138.43, 133.61, 125.36 (*J*_{CF} = 9.8 Hz), 124.13, 115.23 (*J*_{CF} = 24.4 Hz), 112.84 (*J*_{CF} = 19.9 Hz), 35.23, 29.59, 23.03, 15.14; IR (neat) 3046, 2963, 2925, 1623, 1513, 1216, 1136, 817, 767; MS *m/e* 202 (M + 1), 201 (M), 200 (M - 1), 186 (base peak); HRMS for C₁₃H₁₂FN calcd 201.0954, found 201.0954.

8-Fluoro-9-methyl-2,3-dihydro-1*H*-cyclopenta[*b*]quinoline (18b): ¹H NMR (300 MHz, CDCl₃) δ 7.78 (1 H, d, *J* = 8.4 Hz), 7.49 (1 H, td, *J* = 8.0, 5.7 Hz), 7.10 (1 H, dd, *J* = 12.7, 7.8 Hz), 3.16 (2 H, t, *J* = 7.7 Hz), 3.06 (2 H, t, *J* = 7.4 Hz), 2.73 (3 H, d, *J* = 6.2 Hz) 2.19 (2 H, qt, *J* = 7.6 Hz); ¹³C NMR (125 MHz; CDCl₃) δ 167.82, 159.97 (*J*_{CF} = 254.8 Hz), 149.78, 137.44, 135.47, 127.52 (*J*_{CF} = 10.1 Hz), 125.29, 17.91 (*J*_{CF} = 12.2 Hz), 110.62 (*J*_{CF} = 22.9 Hz), 35.18, 29.81, 22.72, 18.71 (*J*_{CF} = 11.9 Hz); IR (neat) 3049, 2961, 1572, 1839, 1221, 1167, 1024, 814, 756 cm⁻¹; MS *m/e* 202 (M + 1), 201 (M), 200 (M - 1), 186 (base peak); HRMS for C₁₃H₁₂FN calcd 201.0954, found 201.0955.

5-Fluoro-9-methyl-2,3-dihydro-1*H*-cyclopenta[*b*]quinoline (19b): ¹H NMR (300 MHz, CDCl₃) δ 7.71 (1 H, d, *J* = 8.3 Hz), 7.40 (1 H, td, *J* = 8.0, 5.3 Hz), 7.31 (1 H, ddd, *J* = 10.7, 7.8, 1.0 Hz), 3.22 (2 H, t, *J* = 7.8 Hz), 3.08 (2 H, t, *J* = 7.4 Hz), 2.59 (3 H, s), 2.21 (2 H, qt, *J* = 7.6 Hz); ¹³C NMR (125 MHz, CDCl₃) δ 167.53, 158.31 (*J*_{CF} = 253.8 Hz), 138.24, 137.70 (*J*_{CF} = 11.4 Hz), 135.31, 129.06 (*J*_{CF} = 3.4 Hz), 124.79 (*J*_{CF} = 8.1 Hz), 119.14 (*J*_{CF} = 3.6 Hz), 112.33 (*J*_{CF} = 19.6 Hz), 35.37, 29.80, 23.03, 15.35; IR (neat) 3044, 2974, 1517, 1194, 814, 760 cm⁻¹; MS *m/e* 202 (M + 1), 201 (M), 200 (M - 1), 186 (base peak); HRMS for C₁₃H₁₂FN calcd 201.0954, found 201.0959.

7-Fluoro-9-phenyl-2,3-dihydro-1*H*-cyclopenta[*b*]quinoline (12d): ¹H NMR (300 MHz, CDCl₃) δ 8.03 (1 H, dd, J = 9.2, 5.6 Hz), 7.54-7.42 (3 H, m), 7.39-7.30 (3 H, m), 7.23 (1 H, dd, J = 9.4, 2.8 Hz), 3.20 (2 H, t, J = 7.6 Hz), 2.89 (2 H, t, J = 7.4 Hz), 2.14 (2 H, qt, J = 7.5 Hz); ¹³C NMR (75 MHz, CDCl₃) δ 166.88, 160.15 ($J_{CF} = 246.3$ Hz), 144.99, 142.11 ($J_{CF} = 4.5$ Hz), 136.26, 134.48, 130.97 ($J_{CF} = 8.3$ Hz), 129.11 (2 C), 128.69 (2 C), 128.21, 127.01 ($J_{CF} = 9.7$ Hz), 117.98 ($J_{CF} = 24.6$ Hz), 109.17 ($J_{CF} = 22.3$ Hz), 35.01, 30.36, 23.49; IR (neat) 3069, 2967, 1629, 1511, 1497, 1217, 833, 708 cm⁻¹; MS *m/e* 264 (M + 1), 263 (M, base peak), 262 (M - 1), 235, 286; HRMS for C₁₈H₁₄FN calcd 263.1110, found 263.1096.

6-Fluoro-9-phenyl-2,3-dihydro-1*H*-cyclopenta[*b*]quinoline (13d): ¹H NMR (500 MHz, CDCl₃) δ 7.74 (1 H, dd, *J* = 8.3, 2.4 Hz), 7.62 (1 H, dd, *J* = 9.2, 6.1 Hz), 7.55-7.47 (3 H, m), 7.35 (2 H, d, *J* = 7.6 Hz), 7.17 (1 H, td, *J* = 8.6, 2.4 Hz), 3.26 (2 H, t, *J* = 7.6 Hz), 2.90 (2 H, t, *J* = 7.4 Hz), 2.18 (2 H, qt, *J* = 7.5 Hz); ¹³C NMR (125 MHz, CDCl₃) δ

168.89, 162.49 (J_{CF} = 247.8 Hz), 149.09 (J_{CF} = 10.9 Hz), 142.90, 136.54, 133.19 (J_{CF} = 3.2 Hz), 129.26, (2 C), 128.66 (2 C), 128.24, 127.75 (J_{CF} = 10.1 Hz), 123.31, 115.51 (J_{CF} = 24.6 Hz), 112.61 (J_{CF} = 20.0 Hz), 35.35, 30.26, 23.58; IR (neat) 3047, 2919, 1574, 1208, 862, 818, 756 cm⁻¹; MS m/e 264 (M + 1), 263 (M, base peak), 262 (M - 1), 235, 186; HRMS for C₁₈H₁₄FN calcd 263.1110, found 263.1090.

7-Methoxy-2,3-dihydro-1 \hat{H} -cyclopenta[b]quinoline (15a): ¹H NMR (300 MHz, CDCl₃) δ 7.89 (1 H, d, J = 9.1 Hz), 7.76 (1 H, s), 7.26 (1 H, dd, J = 9.1, 2.8 Hz), 7.00 (1 H, d, J = 2.8 Hz), 3.89 (3 H, S), 3.11 (2 H, t, J = 7.6 Hz), 3.04 (2 H, t, J = 7.4 Hz), 2.18 (qt, J = 7.5 Hz); ¹³C NMR (75 MHz, CDCl₃) δ 165.34, 157.06, 143.44, 135.87, 129.85, 129.27, 128.24, 120.42, 105.56, 55.42, 34.32, 30.54, 23.65; IR (neat) 3073, 2967, 1507, 1226, 1034, 908, 830 cm⁻¹; MS m/e 200 (M + 1), 199 (M, base peak), 198 (M - 1), 184, 156, 128; HRMS for C₁₃H₁₃INO calcd 199.0997, found 199.0997.

6-Methoxy-2,3-dihydro-1*H*-cyclopenta[*b*]quinoline (16a): ¹H NMR (300 MHz, CDCl₃) δ 7.82 (1 H, s), 7.61 (1 H, d, *J* = 8.9 Hz), 7.37 (1 H, d, *J* = 2.4 Hz), 7.11 (1 H, dd, *J* = 8.9, 2.4 Hz), 3.93 (3 H, s), 3.13 (2 H, t, *J* = 7.6 Hz), 3.06 (2 H, t, *J* = 7.4 Hz), 2.20 (2 H, qt, *J* = 7.5 Hz); ¹³C NMR (75 MHz, CDCl₃) δ 167.83, 160.17, 148.71, 133.50, 130.72, 128.43, 122.51, 118.51, 106.81, 55.51, 34.59, 30.42, 23.71; IR (neat) 2953, 1621, 1415, 1231, 1156, 1029 cm⁻¹; MS *m/e* 200 (M + 1), 199 (M, base peak), 198 (M - 1), 184, 168; HRMS for C₁₃H₁₃NO calcd 199.0997, found 199.0983.

7-Methoxy-9-methyl-2,3-dihydro-1*H*-cyclopenta[*b*]quinoline (15b): ¹H NMR (300 MHz, CDCl₃) δ 7.90 (1 H, d, J = 9.1 Hz), 7.26 (1 H, dd, J = 9.1, 2.7 Hz), 7.15 (1 H, d, J = 2.7 Hz), 3.92 (3 H, s), 3.12 (2 H, t, J = 7.7 Hz), 3.02 (2 H, t, J = 7.4 Hz), 2.51 (3 H, s), 2.17 (2 H, qt, J = 7.5 Hz); ¹³C NMR (75 MHz, CDCl₃) δ 164.53, 157.03, 143.34, 136.81, 134.32, 130.60, 127.88, 119.60, 102.36, 55.52, 34.91, 29.83, 23.10, 15.11; IR (neat) 2982, 1508, 1226, 1022, 832 cm⁻¹; MS *m/e* 214 (M + 1), 213 (M, base peak), 212 (M - 1), 198, 170; HRMS for C₁₄-H₁₅NO calcd 213.1154, found 213.1154.

6-Methoxy-9-methyl-2,3-dihydro-1*H*-cyclopenta[*b*]quinoline (16b): ¹H NMR (300 MHz, CDCl₃) δ 7.82 (1 H, d, *J* = 9.1 Hz), 7.37 (1 H, d, *J* = 2.6 Hz), 7.13 (1 H, dd, *J* = 9.1, 2.6 Hz), 3.93 (3 H, s), 3.13 (2 H, t, *J* = 7.8 Hz), 3.02 (2 H, t, *J* = 7.4 Hz), 2.55 (3 H, s), 2.19 (2 H, qt, *J* = 7.6 Hz); ¹³C NMR (75 MHz, CDCl₃) δ 167.12, 159.71, 148.94, 138.59, 132.09, 124.51, 122.02, 117.79, 107.45, 55.44, 35.13, 29.05, 23.03, 15.01; IR (neat) 2951, 1605, 1507, 1419, 1221, 1153, 1019 cm⁻¹; MS *m/e* 214 (M + 1), 213 (M, base peak), 212 (M - 1), 198; HRMS for C₁₄H₁₅NO calcd 213.1154, found 213.1144.

7-Methoxy-9-phenyl-2,3-dihydro-1*H*-cyclopenta[*b*]quinoline (15d): ¹H NMR (300 MHz, CDCl₃) δ 7.97 (1 H, d, *J* = 9.2 Hz), 7.55–7.43 (3 H, m), 7.38–7.36 (2 H, m), 7.28 (1 H, dd, *J* = 9.2, 2.8 Hz), 6.92 (1 H, d, *J* = 2.8 Hz), 3.72 (3 H, s), 3.19 (2 H, t, *J* = 7.6 Hz), 2.87 (2 H, t, *J* = 7.4 Hz), 2.14 (2 H, qt, *J* = 7.5 Hz); ¹³C NMR (75 MHz, CDCl₃) δ 164.95, 157.19, 143.90, 141.73, 137.03, 134.03, 130.17, 129.20 (2 C), 128.66 (2 C), 128.01, 127.10, 119.96, 104.46, 55.41, 34.94, 30.48, 23.62; IR (neat) 2971, 1620, 1497, 1225, 1022 cm⁻¹; MS *m/e* 276 (M + 1), 275 (M, base peak), 274 (M – 1), 260, 244, 198; HRMS for C₁₉H₁₇NO calcd 275.1310, found 275.1310.

6-Methoxy-9-phenyl-2,3-dihydro-1*H*-cyclopenta[*b*]quinoline (16d): ¹H NMR (300 MHz, CDCl₃) δ 7.54–7.42 (5 H, m), 7.37–7.34 (2 H, m), 7.03 (1 H, dd, J = 9.2, 2.6 Hz), 3.94 (3 H, s), 3.20 (2 H, t, J = 7.6 Hz), 2.88 (2 H, t, J = 7.4 Hz), 2.15 (2 H, qt, J = 7.5 Hz); ¹³C NMR (75 MHz, CDCl₃) δ 167.73, 159.88, 149.76, 142.88, 136.99, 131.55, 129.32 (2 C), 128.54 (2 C), 128.00, 126.78, 121.20, 118.07, 107.39, 55.50, 35.34, 30.23, 23.60; IR (neat) 2965, 1622, 1414, 1225, 1032, 733 cm⁻¹; MS *m/e* 276 (M + 1), 275 (M, base peak), 274 (M – 1), 260, 198; HRMS for C₁₉H₁₇NO calcd 275.1310, found 275.1310.

6-Fluoro-9-(4'-fluorophenyl)-2,3-dihydro-1*H*-cyclopenta[*b*]quinoline (31): ¹H NMR (300 MHz, CDCl₃) δ 7.67 (1 H, dd, *J* = 10.2, 2.6 Hz), 7.56 (1 H, dd, *J* = 9.2, 6.1 Hz), 7.34–7.28 (2 H, m), 7.21 (2 H, t, *J* = 8.8 Hz), 7.14 (1 H, td, *J* = 8.9, 2.6 Hz), 3.20 (2 H, t, *J* = 7.7 Hz), 2.86 (2 H, t, *J* = 7.4 Hz), 2.16 (2 H, qt, *J* = 7.5 Hz); ¹³C NMR (75 MHz, CDCl₃) δ 168.84, 162.59 (*J*_{CF} = 249.3 Hz), 162.46 (*J*_{CF} = 249.2 Hz), 149.02 (*J*_{CF} = 12.2 Hz), 141.82, 133.33, 132.34 (*J*_{CF} = 5.1 Hz), 130.99 (2 C, *J*_{CF} = 9.8 Hz), 127.39 (*J*_{CF} = 9.7 Hz), 123.24, 115.77 (2 C, *J*_{CF} = 22.0 Hz), 115.62 (*J*_{CF} = 24.4 Hz), 112.66 (*J*_{CF} = 21.8 Hz), 35.26, 30.22, 23.52; IR (neat) 3051, 2963, 1607, 1499, 1211, 839 cm⁻¹; MS *m*/*e* 282 (M + 1), 281 (M, base peak), 280 (M - 1), 186; HRMS for C₁₈-H₁₃F₂N calcd 281.1016, found 281.1027.

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Registry No. 8a, 2468-55-5; **8b**, 28077-74-9; **8c**, 132017-60-8; **8d**, 34886-50-5; **9**, 931-54-4; **10a**, 5661-06-3; **10b**, 6829-07-8; **10c**, 132017-61-9; **10d**, 10265-82-4; **11**, 24075-34-1; **12a**, 1717-19-7; **12b**, 132017-

62-0; 12d, 132017-63-1; 13a, 132017-64-2; 13b, 132017-65-3; 13d, 132017-66-4; 14, 10349-38-9; 15a, 61831-48-9; 15b, 132017-67-5; 15d, 132017-68-6; 16a, 30160-18-0; 16b, 132017-69-7; 16d, 132017-70-0; 17, 24075-35-2; 18a, 132017-71-1; 18b, 132017-72-2; 19a, 132017-73-3; 19b, 132017-74-4; 30a, 132017-75-5; 30a (debromo derivative), 14636-27-2; 30b, 132017-76-6; 30b (debromo derivative), 132017-78-8; 31, 132017-77-7; PhCH=CH(CH₂)₃I, 132017-79-9; 2-benzylcyclopentanone, 2867-63-2.

Absorbance, Light Intensity, Mass Transfer, and Sampling Time Effects in a Proposed Mechanism for the Photolysis of Phenyl Azide

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Abstract: In the concentrated phenyl azide solutions in which Costantino et al. have reported apparent quantum yields for azide disappearance ($\phi_{-phN_{2}}$) on the order of 1000 and interpreted their results in terms of a branching-chain mechanism, the optical penetration depth is sufficiently small that the proposed chain-initiation reaction is effectively confined to a very thin layer near the front optical window. Simple calculations show that this layer is sufficiently thin that diffusive mass transfer is important on the time scale of the experiments. On the basis of the mechanism of Costantino et al., we develop and investigate a model consisting of three nonlinear integro-partial differential equations accounting for the nonuniform light absorption, photochemical kinetics, and diffusive mass transfer. The dependence of ϕ_{-PhN_3} on initial azide concentration is shown to be in good qualitative agreement with experiment. The model predicts that the apparent quantum yield is sometimes a strong function of the time at which the reaction mixture is sampled. The relatively simple mechanism proposed by Costantino et al. is consistent with their results, as well as the more recent measurements of Liang and Schuster, in which quantum yields in excess of unity were not observed. In terms of the proposed branching-chain mechanism, the apparent discrepancies between the results of Costantino et al. and Liang and Schuster can be attributed to the use of different light intensities.

In large part due to their use as photolabeling agents, polymerization initiators, and cross-linking agents,¹ there has been considerable recent interest in the photochemistry of aryl azides,²⁻⁹ with special emphasis on the photolysis of phenyl azide. Recently, additional interest has developed in this area due to the fact that the photolysis can lead to the formation of poly(1,2-azepines). When the process is performed through a photomask, the oxidation products of the poly(1,2-azepines) can form a patterned, semiconducting polymeric film¹⁰ of interest in microelectronics manufacture.

In dilute solution, the photodecomposition of phenyl azide at 254 nm proceeds with a quantum yield (for the disappearance of phenyl azide) of about 0.5, in both nonpolar ($\phi_{N_2} = 0.53 \pm 0.10$ in hexane at 25 °C and 0.52 \pm 0.10 in a matrix of methylcyclohexane at 77 K)¹¹ and polar ($\phi_{-PhN_3} = 0.57 \pm 0.19$ in deoxygenated acetonitrile)² solvents.

Recently, Costantino et al.² have reported that, in the photolysis of concentrated (>10⁻³ M) solutions of phenyl azide in acetonitrile, ϕ_{-PhN_3} may exceed 10⁴. They have proposed a branching-chain mechanism involving triplet nitrene radicals, a chain-initiation step (1), a chain-branching step (2), and two chain-termination steps (3 and 4), which is proposed to be valid over the entire range of azide concentration. Jenkins et al.³ have conducted a mech-

$$PhN_3 \xrightarrow{n\nu} PhN + N_2$$
 (1)

$$PhN + PhN_3 \rightarrow [X] \rightarrow 2PhN + N_2 \tag{2}$$

$$PhN + PhN_3 \rightarrow Ph - N = N - Ph + N_2$$
(3)

$$2PhN \rightarrow Ph-N=N-Ph \qquad (4)$$

anistic study of the photodecomposition of several substituted phenyl azides in the presence of various triplet sensitizers as well as a known singlet quencher and have concluded that the intermediate nitrene in (1-4) is a triplet species.

Recently, Liang and Schuster⁹ and Li et al.¹² have questioned the validity of the mechanism (1-4) at high azide concentrations. Using time-resolved infrared and ultraviolet absorption spectroscopy to monitor the concentration of 1,2-didehydroazepine following flash photolysis of phenyl azide, Li et al. have proposed an alternative mechanism, in which the singlet excited azide undergoes intersystem crossing to an excited triplet azide or loses N_2 to form a singlet nitrene. The latter species may undergo either a reversible isomerization to 1,2-didehydroazepine or intersystem crossing to a triplet nitrene, a species which may also result from loss of N_2 by triplet azide. Li et al. propose that azobenzene is formed by dimerization of two triplet nitrenes and that 1,2-didehydroazepine either reacts with azide to give a (presumably polymeric) tarry substance or is converted to triplet nitrene by a process not involving isomerization to singlet nitrene. At low

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